perience it is easy to reach a purity degree of at least 90%, in most cases 98%. Until now, a comparable purity and recovery was only obtained from blood of patients having an eosinophilia of 40-85%. In these cases the method of Day11 gives good results, but it can not be used for the isolation of the eosinophils from normal human blood.

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Other special and more complicated methods published recently (Spray<sup>12</sup>, Shar<sup>13</sup> and Parrillo<sup>14</sup>) use the differences in IgG binding capacity, the different iron phagocytosis or differences in the adherence on a nylon wool surface of the neutrophilic and eosinophilic granulocytes, for isolation of the eosinophils from normal blood.

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## A simple and versatile apparatus for the continuous superfusion of nervous tissue preparations

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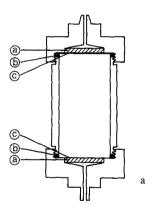
Department of Medical Biochemistry, University of the Witwatersrand Medical School, Johannesburg 2001 (South Africa), 2 Āpril 1979

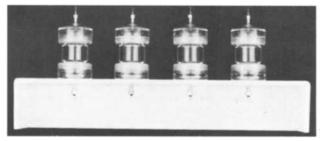
Summary. A bimodal apparatus for the continuous superfusion of both subcellular particles and whole tissue slices is described.

The measurement of the rate of release of neurotransmitter substances from slices or subcellular fractions of nervous tissue has been confounded by the presence of very active transport systems. These uptake processes, which exist for many transmitters in nerve endings, synapse-associated glial cells and post-synaptic perikarya, are responsible for the removal of the released substance from the surrounding medium before an accurate measurement of release can be made. Recently, this problem has been overcome by the development of a superfusion apparatus shown to minimise reuptake of released transmitter by continuous removal of the medium and released transmitter substance<sup>2,3</sup>. In this communication, we describe a simple apparatus which can be used for the continuous superfusion, at constant volume, of subcellular particles or whole brain slices.

The chamber depicted in the figure consists of a central barrel of machined perspex which is screwed into 2 identical perspex endpieces. Each endpiece is fitted with a Millipore 25 mm filter support frit and a rubber 'O' ring (Millipore) to seal the connection. The outlet of the endpiece is machined to give minimum dead volume. The apparatus can be used in the conventional manner to superfuse a bed of subcellular particles resting on the lower Millipore filter. In this mode, an appropriate volume of medium can be introduced into the chamber and the volume thereafter kept constant since the apparatus is airtight. It is also possible to superfuse subcellular organelles or whole tissue slices against gravity by pumping the medium into the chamber from the bottom of the apparatus and drawing it out of the top. Such upward displacement of the medium keeps the tissue preparation in suspension and prevents blockage of the lower filter, a problem often encountered in superfusion of tissue slices by conventional techniques. Since it is necessary to fill the chamber completely with medium when operating in this mode, the volume of the chamber can be varied by altering the length of the barrel.

An apparatus consisting of 4 chambers of the type described above arranged in parallel has been used extensively in the investigation of the efflux of the amino acid neurotransmitters gamma-aminobutyric acid and L-gluta-





Superfusion apparatus. a Diagram of the construction of the perspex apparatus in cross section. All dimensions are based on the 25 mm Millipore filter shown at (c). Rubber 'O' rings are shown (b) together with the Millipore support frits (a). b The complete apparatus mounted in a bank of 4 chambers as used in experiments to determine neurotransmitter release.

mate from nervous tissue preparations under a variety of conditions<sup>3–5</sup>. In addition, we have used the apparatus to monitor the release of endogenous somatostatin from tissue slices and synaptosomes (unpublished results).

Since the chambers can be used in 2 different modes and the superfusion volume and flow-rate maintained constant, we believe that this apparatus has wide application in the perfusion of preparations from a variety of tissues. Certainly in the study of neurotransmitter release mechanisms, where uptake has been shown to interfere with measurement of the true rate of efflux, this design has proved both versatile and invaluable.

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